PROTEIN KINASE C IOTA IS REQUIRED FOR PANCREATIC CANCER CELL MAYO CLINIC **TRANSFORMED GROWTH AND TUMORIGENESIS Cancer Center**

ABSTRACT:

Pancreatic cancer is the fourth leading cause of cancer deaths in the United States with an overall 5-year survival rate of <5%. Pancreatic ductal adenocarcinoma (PDAC), the most common form of pancreatic cancer, is highly resistant to conventional chemotherapies; therefore, there is a critical need for new molecular targets for pancreatic cancer chemotherapy. Mutational activation of the KRAS proto-oncogene occurs in >90% of PDAC. Oncogenic K-ras activates atypical protein kinase C iota (PKCi) and PKCi is required for oncogenic Ras-mediated transformed growth in lung cancer and intestinal epithelial cells. However, little is known about the role of PKCi in pancreatic cancer. In this study we evaluated the requirement for PKC₁ for the transformed growth and tumorigenicity of PDAC cells. Our results demonstrate that PKC₁ is significantly over-expressed in human pancreatic cancer and is required for PDAC cellular transformation in vitro and in vivo. Specifically, inhibition of PKC₁ expression blocks PDAC cell transformed growth in vitro and tumorigenicity in vivo. Analysis of PKCi downstream effectors implicates Rac1/(PAK)-MEK-ERK1/2 signaling in PKCimediated transformed growth. Inhibition of PKC₁ expression in orthotopic pancreatic tumors also significantly reduces tumor angiogenesis and metastasis. Taken together, our data demonstrate a required role for PKC₁ in the transformed growth of pancreatic cancer cells and documents a novel role for PKC₁ in pancreatic cancer cell metastasis and angiogenesis *in vivo*. These results strongly suggest that PKC₁ will be an effective target for pancreatic cancer therapy.



Figure 1: PKC_l is highly expressed in human pancreatic cancer and PDAC cell lines.

A) qPCR analysis of PKC₁ mRNA expression in 28 matched human pancreatic tumor and adjacent non-tumor pancreas. mRNA abundance is normalized to 18S x 104. Horizontal line indicates the mean PKC₁ mRNA abundance in non-tumor matched tissues + 2*(SD). Inset: PKC₁ mRNA expression is significantly increased in tumors compared to normal pancreas tissue. The mean PKC₁ mRNA abundance in non-tumor matched tissues is set equivalent to 1, and average fold-increase is plotted for pancreatic tumors. B) A representative image of immunohistochemical detection of PKC expression in a formalin-fixed primary pancreatic cancer reveals a high level of expression compared to normal adjacent human pancreas (bottom, right panel). C) qPCR analysis of PKC₁ mRNA expression in ten human pancreatic cancer cell lines (top). mRNA abundance is normalized to GAPDH x 100, n=3. Immunoblot analysis of total cell lysates from ten human pancreatic cancer cell lines for expression of PKC_{ι} and β -actin (bottom)



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Figure 2: PKC_l is not required for anchorage-dependent (nontransformed) growth of PDAC

qPCR analysis of PKCi mRNA expression in A) Panc-1 and B) MiaPaCa-2 stably carrying either non target (NT), PKC₁-specific RNAi constructs (PKC₁ #1) or (PKC₁ #2). Analysis was performed in triplicate and is representative of two independent experiments. Insets, Immunoblot analysis of PKC_i, PKC ζ and β -actin protein expression in A) Panc-1 and B) MiaPaCa-2 NT or PKC₁-RNAi (PKCi#1 and PKCi#2) constructs. Anchorage-dependent growth in C) Panc-1 and D) MiaPaCa-2 stably carrying either NT or PKC₁-RNAi (PKC₁#1 and PKC₁#2) was determined by MTT colorimetric





Figure 4: Constitutively active Rac1 recovers transformed growth of PKC_l RNAi PDAC cells. A) Panc-1 cells stably expressing NT or PKC_l RNAi were assayed for Rac1 activity. Top panel, (Active) Rac1-GTP was precipitated from cell extracts with PAK-1 PBD agarose. Immunoblot analysis of precipitates and total cellular extracts (total Rac1) was performed using an anti-Rac1 Ab. Bottom panel, Quantitative, densitometric analysis of relative Rac1 activity (active Rac1/total Rac1). Mean of three independent experiments +/- SEM is plotted. B) Panc-1 cells cotransfected with RNAi (NT or PKC_i) and control vector (LZRS) or vector expressing constitutively active (ca) Rac1 (caRac) were subject to immunoblot analysis for expression of Rac1, PKCi, phospho-ERK1/2 (Thr202/Tyr204), ERK1/2 and actin as a loading control. C) Expression of caRac1 recovers the inhibitory effect of PKC₁ RNAi on soft agar colony formation. *= significantly different than control (NT & LZRS), **= significantly different than PKC_l RNAi & LZRS. Mean +/- SEM is plotted and represents at least two independent experiments.



Figure 5: Inhibition of PKC_l blocks orthotopic pancreatic tumor proliferation and proliferative signaling. A) Tumor growth was monitored by bioluminescence (total flux, photons/sec) detected by IVIS imaging of orthotopic Panc-1 NT versus Panc-1 PKC₁ RNAi pancreatic tumors in live, anesthetized mice at weekly intervals after tumor implantation. *= significantly different than NT RNAi tumors. n=15-16/group. B) Top: Immunohistochemical analysis of BrdU incorporation. Bar=100µm. Bottom: Quantitative analysis of BrdU incorporation into Panc-1 tumors. Mean +/-SEM is plotted. C) Immunohistochemical detection of PKC₁ and phospho-ERK1/2 (Thr 202/ Tyr 204) (representative images). Bar=100µm. D) Representative immunoblot analysis of PKCi, phospho-ERK1/2 (Thr 202/ Tyr 204) and ERK1/2 in Panc-1 NT and PKC₁ RNAi orthotopic pancreatic tumors. Equivalent amounts of protein from each tumor sample were analyzed.

Figure 3: PKC_l is required for anchorage-independent growth of PDAC cells.

Soft agar colony formation of A) Panc-1 and B) MiaPaCa-2 cells with NT or PKC₁-RNAi (PKCi#1 and PKCi#2) constructs. *= significantly different than NT. C) Immunoblot analysis of PKC₁ expression in Panc-1 cells co-transfected with RNAi (NT or PKC_l) and control vector (pBabe) or vector expressing wild type PKCı (PKC_l). D) Re-expression of WT PKC_l overcomes the inhibitory effect of PKCu RNAi on soft agar colony formation. *= significantly different than control (NT & pBabe), **= significantly different than PKC₁ RNAi & pBabe. Mean +/- SEM is plotted and represents at least two independent experiments.



Figure 6: Inhibition of PKC_l blocks PDAC angiogenesis and metastasis. A) Left: Immunohistochemical detection of CD31 staining. Right: Quantitative analysis of CD31 positive staining in Panc-1 tumors, calculated as the ratio of CD31-positive pixels to the sum of all pixels. Mean +/- SEM is plotted. Bar=100µm. B) Representative immunoblot analysis of VEGF and actin in Panc-1 NT and PKC₁ RNAi orthotopic pancreatic tumors. Equivalent amounts of protein from each tumor sample were analyzed. C) Bioluminescence IVIS images of orthotopic Panc-1 NT versus PKC₁ RNAi pancreatic tumors in live, anesthetized mice at Day 35. D) Representative images of tumor metastases to various organs. E) Percent of orthotopic Panc-1 NT and PKC₁ RNAi pancreatic tumors that metastasized to various organs is plotted. *= significantly different than NT RNAi tumors.

CONCLUSIONS:

 \geq PKC₁ expression is highly overexpressed in a high percent of primary pancreatic cancer tumors and highly induced in PDAC cell lines.

 \geq PKC_l is dispensable for adherent pancreatic cell growth, but is required for transformed growth of pancreatic carcinoma cells in vitro.

 \geq PKC₁ and its downstream effector Rac1 are required for PDAC transformed growth *in vitro*.

 \geq In vivo, PKC_i regulates PDAC tumorigenicity and tumor cell proliferation. >PKCι expression regulates ERK activation in vivo, suggesting a Rac1-MEK-ERK signaling pathway is involved in PKCιdependent PDAC tumor cell proliferation in vivo.

>Inhibition of PKC_l expression in PDAC cells reduces PDAC tumor angiogenesis and metastasis.

Our results demonstrate a requirement for PKC₁ in transformed growth and oncogenic signaling in pancreatic cancer cells in vitro and in vivo. Taken together, these data strongly implicate PKCL as a candidate therapeutic target for the treatment of pancreatic cancer.

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