

# Structure/function analysis of TDP-43 neurotoxicity in *C. elegans*

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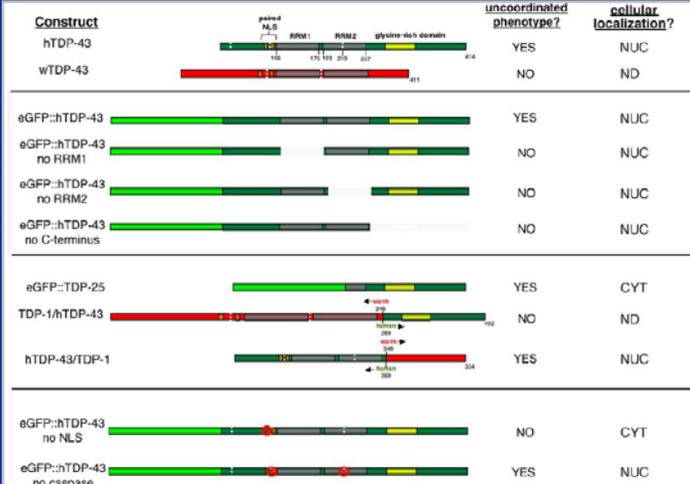
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## ABSTRACT

TDP-43 is a conserved RNA binding protein with known roles in mRNA splicing and stability. Cytoplasmic deposition of TDP-43 has been linked to multiple neurodegenerative diseases, including ALS and frontotemporal lobar dementia (FTLD). We have engineered pan-neuronal expression of human TDP-43 protein in *C. elegans*, with the goal of generating a convenient *in vivo* model of TDP-43 neurotoxicity. Full-length (wild type) human TDP-43 expressed in *C. elegans* is nuclear as is observed in human cells. Transgenic worms with neuronal human TDP-43 expression exhibit an uncoordinated phenotype and have abnormal motoneuron synapses. By using this uncoordinated phenotype as a read-out of TDP-43 neurotoxicity, we have investigated the contribution of specific TDP-43 domains as well as TDP-43 sub-cellular localization to toxicity. Deletion of either RNA recognition domain (RRM1 or RRM2) completely blocks neurotoxicity, as does deletion of the C-terminal region. These deleted TDP-43 variants still accumulate in the nucleus, although their subnuclear distribution is altered. In contrast, N-terminal deletions result in the formation of toxic cytoplasmic aggregates. Mutation of the TDP-43 nuclear localization signal (NLS) results in cytoplasmic deposition of full-length TDP-43, which is not toxic. Mutations that alter two TDP-43 caspase cleavage sites (D89/219E), however, do not reverse TDP-43 toxicity. Our results demonstrate that TDP-43 neurotoxicity can result from either nuclear activity of the full-length protein or accumulation of cytoplasmic aggregates composed of C-terminal fragments. These results suggest that there may be (at least) two different mechanisms of TDP-43 neurotoxicity.



**Figure 2:** Summary of temperature inducible *snb-1* driven constructs expressed in *C. elegans*; their respective binary phenotypic outcome (unc/unc) and their cellular localisation.

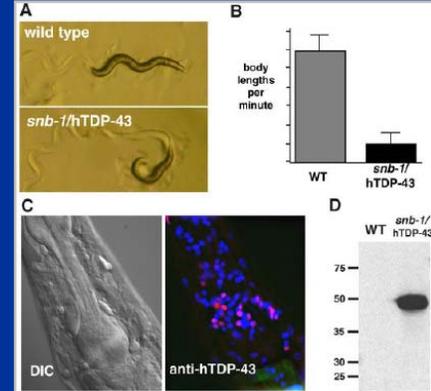
## SUMMARY OF RESULTS

- Pan-neuronal expression of full length nuclear hTDP-43 in *C. elegans* produces uncoordinated movement.
- GABAergic motoneuron synaptic dysregulation and axonal fasciculation is observed, but not motoneuron loss.
- This phenotype is alleviated by deletion of the functional domains RRM1, RRM2 and the C terminal domain.
- This phenotype is alleviated by mutagenesis of the hTDP-43 NLS.
- The unc phenotype is recapitulated by pan-neuronal cytoplasmic expression of the ALS relevant C terminal fragment TDP-25.
- The mechanisms of nuclear hTDP-43 and cytoplasmic hTDP-25 derived neurotoxicity can be disseminated from each other using the caspase-cleavage resistant construct hTDP-43.D89E, D219E.

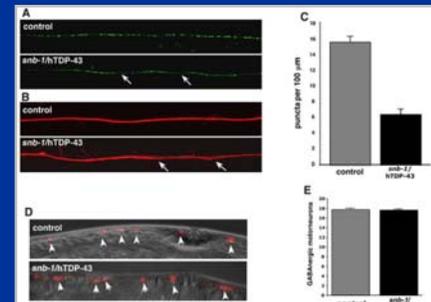
## CONCLUSIONS

We report two mechanisms of hTDP-43 neurotoxicity in *C. elegans*, from:

1. Pan-neuronal expression of full length nuclear hTDP-43.
2. Cytoplasmic aggregation of the ALS relevant C terminal fragment TDP-25.



**Figure 3:** A. Binary phenotypes for wild type and *snb-1/hTDP-43* transgenic worms. B. Quantification of movement defects in *snb-1/hTDP-43* worms (strain CL2029). C. Nerve ring of fixed and permeabilized *snb-1/hTDP-43* worms probed with anti-hTDP-43 antibody (ProteinTech anti-TASBP polyclonal antibody). Red, anti-hTDP-43; blue, DAPI staining of nuclei; green, intestinal GFP from transformation marker plasmid. D. Immunoblot of extracts from wild type and *snb-1/hTDP-43* transgenic worms (strain CL1682) probed with anti-TDP-43 monoclonal antibody M01.



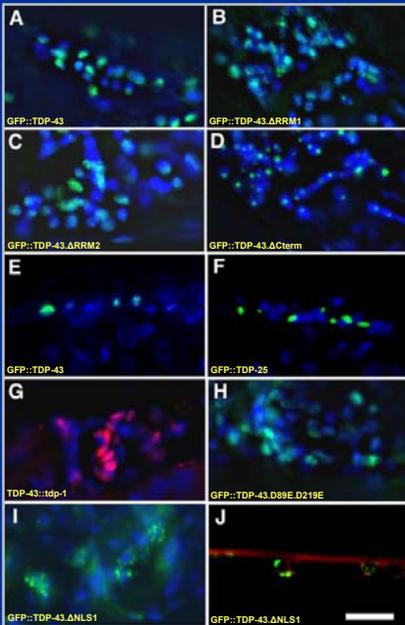
**Figure 4:** Neuropathology in *snb-1/hTDP-43* transgenic worms. A. GABAergic motor neuron synapses in dorsal cord of living control (CL1685) and *snb-1/hTDP-43* (CL1681) worms using an unc-25::SNB-1::GFP reporter transgene (juv1). B. Dorsal cord axonal processes in wild type and *snb-1/hTDP-43* worms, visualized by co-injection with *gfp-1::Daf-2c*, a reporter that accumulates in all axonal processes (32). Note defasciculations in *snb-1/hTDP-43* axonal bundles (arrow). C. Quantification of dorsal GABAergic synapses in control (CL1685) and *snb-1/hTDP-43* (CL1681) worms. *N=30*. D. Visualization of GABAergic motor neurons using *unc-47::Daf-2c* reporter transgene (*hdsf22*). E. Quantification of GABAergic motor neurons in control and *snb-1/hTDP-43* worms. *N=30*.

## REFERENCES

- Ayala YM, et al. (2005) Human, *Drosophila*, and *C. elegans* TDP43: nucleic acid binding properties and splicing regulatory function. *J Mol Biol* 348: 575-88.
- Neumann M, et al. (2006) Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science* 314: 130-3.
- Mackenzie I, et al. (2006) Heterogeneity of ubiquitin pathology in frontotemporal lobar degeneration: classification and relation to clinical phenotype. *Acta Neuropathol*. 112(5): 539-549.
- Zhang Y, et al. (2009) TDP-43 C-terminal fragments enhances neurotoxicity and induces cytoplasmic inclusions *in vivo*. *Proc Natl Acad Sci U S A*. Published online April 21, 2009.

This work was supported by:

Mayo Clinic Foundation (L.P.), the National Institutes of Health/National Institute on Aging [R01AG026251 and P01AG17216-06 (L.P.)], the National Institutes of Health/National Institute of Neurological Disorders and Stroke [R01 NS 063964-01 (L.P.) and R01 NS063964 (C.D.L.)], the Amyotrophic Lateral Sclerosis Association (L.P.) and the Department of Defense USAMRMC PR080354 (L.P.).



**Figure 1:** *snb-1*-driven full-length and variant hTDP-43 constructs. A to J fixed and counterstained with DAPI. A. Nerve ring area of øGFP::hTDP-43 transgenic worm (CL1626). B. Nerve ring of øGFP::hTDP-43 RRM2 deletion (Δaa106 to 175) transgenic worm (CL1702). C. Nerve ring of øGFP::hTDP-43 RRM2 deletion (Δaa193 to 257) transgenic worm (CL1705). D. Nerve ring of øGFP::hTDP-43 C-terminal deletion (hTDP-43 1-257, strain CL1710). E. Ventral cord region of øGFP::hTDP-43 transgenic worm (CL1626). F. Ventral cord region of øGFP::TDP-25 expressing worm (F1 transgenic animal). G. Nerve ring of hTDP-43/TDP-1 fusion construct probed with anti-hTDP-43 monoclonal antibody (red). H. Nerve ring of øGFP::hTDP-43 caspase-cleavage resistant (D89E, D219E) transgenic worm. I. Nerve ring of øGFP::hTDP-43 NLS1 mutant transgenic worm (strain CL1687). J. Ventral cord region of live øGFP::hTDP-43 NLS1 mutant worm (CL1687) containing co-injected *gfp-1::Daf-2c* marker to highlight cytoplasm in axons and cell bodies. Size bar = 5µm.